

**FRESH MADRE DE AGUA (*Trichanthera gigantea*) AND MALUNGGAY (*Moringa oleifera*) LEAVES AS FEED INCLUSION FOR JAPANESE QUAIL (*Coturnix japonica*)**

Melanie L. Tecson<sup>1</sup> and Kent Marcial L. Catubis<sup>2</sup>

**ABSTRACT**

The growth, egg quality and cost-benefit ratio was determined in this study wherein 120 ready-to-lay quail birds were used. The birds were distributed to six treatments, namely: T0 - control ration based on commercial feeds (quail layer mash); T1 - 10g fresh *Trichanthera gigantea* leaves (FTL); T2 - 10g fresh *Moringa oleifera* leaves (FML); T3 - 5g FTL and 5g FML, T4 - 7g FTL and 3g FML and T5 - 3g FTL and 7g FML. The egg weight, eggshell thickness, albumen and yolk weight, breadth and length, of the Japanese quail birds were gathered every two weeks. Furthermore, the body weight was measured every two weeks while feed consumption and number of eggs produced were recorded daily. The average body weight and the number of eggs produced significantly differed on week 8; T0 had the highest body weight and eggs produced, while T2 had the lowest body weight and eggs produced. Moreover, T5 had the thickest eggshells among the treatments. However, there were no significant differences in egg weight, albumen, yolk, breadth, length, and feed consumption. Furthermore, the inclusion of fresh *T. gigantea* and *M. oleifera* leaves of 3g and 7g (T5) positively affected the eggshell thickness. Moreover, it shows T5 show potential return of utilization in production.

Keywords: quail, *Trichanthera*, *Moringa*, egg

**INTRODUCTION**

The correct balance of calories, protein, fat, carbohydrates, vitamins, and minerals provides energy, and the variety of nutrients is what growing children and working adult needs (Tunsaringkarn *et al.*, 2013). Animal protein, especially in the poultry sub-sector, is one of the most important sources of such nutrients needed in human nutrition because of its biological significance (Olorunfemi *et al.*, 2016). The egg is one of the human diet's most common and abundant protein sources. Together with chicken, quail (*Coturnix japonica*) is one of the poultry species which recently gained popularity as a source of animal protein. The essential amino acid content of Japanese quail eggs comprised 50.36% of albumen protein and 48.65% of yolk protein. The polyunsaturated fatty acid (PUFA) content in the phospholipid fraction of the yolk was almost 2.5 times higher than in the triglyceride fraction,

---

<sup>1</sup>Animal Science Department, College of Agriculture, Cebu Technological University – Tuburan Campus, Barangay 8, Tuburan Cebu, Philippines; <sup>2</sup>Crop Science Department, College of Agriculture, Cebu Technological University – Tuburan Campus, Barangay 8, Tuburan Cebu, Philippines (email: melanie.tecson@ctu.edu.ph).

whereas polyunsaturated fatty acid:saturated fatty acid (PUFA:SFA) ratio was 0.52:1 vs 0.26:1-0.28:1 in the triglyceride fraction. Out of albumen mineral substances, the highest content was that of phosphorus, calcium and magnesium. The yolk was the richest in phosphorus and calcium, whereas the eggshell was in calcium (Genchev, 2012).

Moreover, quail eggs has a considerable number of antioxidants, minerals, and vitamins higher than other eggs (Lalwani, 2011). Due to its petite body, the Japanese quail birds only require a smaller house than chickens. The content of the essential amino acid leucine was the highest (1.37%) in the yolk of chicken eggs, comparable to the concentration of this amino acid in the yolk of quail eggs (1.4%) (Tolik *et al.*, 2014). The total antioxidant of quail egg activities on the yolk and albumen extracts was reported at 186.57+6.441 mg/g and 172+10.690 mg/g ascorbic acid, respectively (Oladipo and Ibukun, 2017). The vitamin content of quail egg yolk are fat-soluble wherein vitamin E (tocopherol, 5920.0 µg/100g) was significantly higher than vitamin A (717.0 µg/100g,  $P<0.001$ ) and vitamin D (1.14 µg/100g,  $P<0.001$ ). Furthermore, the essential minerals of whole eggs were nitrogen (6.36%) which was mostly in egg whites (12.2%), while most trace minerals of whole eggs were iron (80.8 mg/Li) and zinc (46.9 mg/Li). Both iron (116.0 mg/Li) and zinc (70.6 mg/Li) were higher in egg yolks (Tunsaringkarn *et al.*, 2013).

Most countries depend on the importation of feeds for most poultry species (including quail), especially on meals rich in protein and energy. This usually resulted in high production costs that ultimately led to an increased market price of poultry products (Bahadori *et al.*, 2017). Feed cost is one of the highest costs in poultry production accounting for 70 – 80% of all components of production costs incurred (Pramono *et al.*, 2018). In solving such a problem, other countries resorted to the research of the use of some organic-based materials as feed inclusion to minimize feed costs without compromising the nutrition quality (Ronald and Adamchak, 2018) In addition, sturdy eggshell indicates good egg quality, thus, calcium-based supplements must be incorporated on quail feeds. The horseradish tree (*Moringa oleifera*), locally known as *Moringa*, has proven to be an abundant and rich source of nutrients, especially vitamins and minerals (Hasan *et al.*, 2019).

The research conducted by Wilson and Hettinger (2017) in the Philippines revealed that 100g or one cup of cooked *Moringa* leaves contain 3.1g of protein, 0.6g of fiber, 96mg of calcium, 29mg of phosphorus, 1.7mg of iron, 2,820mg of beta-carotene, 0.07mg of thiamin, 0.14mg of riboflavin, 1.1mg of niacin and 53mg of ascorbic acid (PCHRD-DOST, 2018). Abbas (2013) reported that *Moringa* leaf meal could be used up to 6% of the diet of growing layer chicks, 10% of laying hen diets and 5% of broiler diets without deleterious effects on their growth and laying performance. Moreover, *T. gigantea* leaf has 65.43% crude protein, 72.73% lipid, 23.08% fiber and 68.90% non-nitrogen extract digestibility, making it a palatable feed inclusion for livestock (Hien *et al.*, 2018). Furthermore, the organic food market is expanding worldwide with the rising concern in food safety, health-conscious individuals and environmental protection (Malkanathi *et al.*, 2021).

Due to the increasing consumer awareness and demand for organic and quality quail meat and egg, this study was conducted using *T. gigantea* and *M. oleifera* leaves as feed inclusion for Japanese quails' growth and laying performance. Moreover, the study examined the egg quality of quails fed with such formulated feeds, which usually needed to be improved in every quail study. Lastly, it also evaluated whether such materials can decrease feed costs for quail producers without compromising the quality.

## MATERIALS AND METHODS

All the procedures in caring for animals in this study were submitted for review and approved by the Institutional Animal Control and Use Committee (IACUC) of Cebu Technological University (CTRU) - Main Campus with PROTOCOL no. IACUC-TU-AO5. After complying with the ethical requirements, the study was guided by the research standard.

The experimental house was constructed at the Cebu Technological University (CTU) – Tuburan Campus livestock farm. The cage size per treatment was 1m x 1m x 0.5m. The cage was made of plywood, ¼ inch mesh wires and 1in x 1in lumber for the framework (Offiong *et al.*, 2020). A linear feeder was provided outside the cage. A corrugated galvanized iron sheet was used for the roofing wherein an artificial insulator was attached.

*T. gigantea* and *M. oleifera* leaves were collected inside the CTU Tuburan Campus Farm. Fresh leaves were chopped and mixed into other components in the respective diets. A completely randomized design (CRD) was used in the study. A total of 120 ready-to-lay Japanese quail birds (approximately 150-200g body weight and six weeks old) were used in the experiment and purchased at the hatchery of Cebu City. The birds were distributed to six (6) treatments with four (4) replicates and five (5) samples per replicate. The treatments used were as follows:

- T0 – 40g (Recommended Commercial feeds only) control
- T1 – 30g Commercial quail layer mash + 10g Fresh *Trichanthera gigantea* leaves (FTL)
- T2 – 30g Commercial quail layer mash + 10g Fresh *Moringa oleifera* leaves (FML)
- T3 – 30g Commercial quail layer mash + 5g FTL + 5g FML
- T4 – 30g Commercial quail layer mash + 7g FTL + 3g FML
- T5 – 30g Commercial quail layer mash + 3g FTL + 7g FML

The Japanese quails were acclimatized for two (2) weeks prior to the administration of treatments that lasted for two (2) months. The birds were given electrolytes for three (3) days to alleviate transportation stress. Water was provided *ad libitum* throughout the experiment. Furthermore, other factors affecting the quail birds were given appropriate attention and action.

Data gathered were as follows:

### 1. Growth Performance

Each quail's initial body weight was collected a day before administering the treatments. Weight gain was gathered every two (2) weeks until the end of the study using a digital weighing scale.

### 2. Egg Parameters

The eggs produced per day were recorded. Measurement of shell thickness (mm), egg length (mm) and egg width (mm) were measured through the Mitutoyo 500-171-30 AOS Absolute Caliper 6"/ 150mm SPC. While egg weight (g) was measured using a digital weighing scale. Lastly, average egg weight (g) was calculated weekly, while other parameters were evaluated monthly until the termination of the experiment.

### 3. Feed consumption

Feed intake (g) and average daily gains (ADG) were computed using the formulas below:

$$\text{Feed Intake} = \text{Feeds Offered} - \text{Feed Refusal}$$

$$\text{Average Daily Gain} = \frac{\text{Total weight gain}}{\text{Number of days between start weight and Current Weight}}$$

### 4. Feed Proximate Composition Analysis

Each feed treatment sample was submitted to the Department of Science and Technology – Region 7 and Fast Laboratories for proximate composition analysis (crude protein, ash content, crude fiber, moisture, calcium).

### 5. Cost-Benefit Ratio

Cost-Benefit-Ratio was computed by the future expenses and income based on the two (2) months of study. This was calculated by dividing the future income by expenses for five (5) years using the formula below:

$$\text{Benefit-Cost Ratio} = \frac{\sum \text{Present Value of Future Benefits}}{\sum \text{Present Value of Future}}$$

All the data were consolidated and transferred to MS Excel for tabulation. The Analysis of Variance (ANOVA) was used to analyze significant differences among treatments, and differences among treatment means were analyzed using Tukey. Correlation among variables was analyzed using the Minitab Statistical Software.

## RESULTS AND DISCUSSION

The quality and performance of the farm animals matters on the components of nutrients found in feeds. The percentage of the proximate analysis shown in Table 1 was varied, which implies that the said treatment had different compositions of nutrients.

However, the egg numbers were significantly different among treatments, as shown in Table 2. The control group had the highest while T2 with 10g of fresh *M. oleifera* leaf had

Table 1. Proximate Analysis of *Trichanthera gigantea* and *Moringa oleifera* leaves in the treatment diets.

Nutrient	T1	T2	T3	T4	T5
Crude Protein	16.00	12.60	8.47	9.59	10.30
Crude Fiber	7.04	4.48	5.86	6.98	7.06
Crude Fat	3.86	4.82	4.21	3.84	3.99
Ash	15.95	10.03	13.30	11.28	12.01
Calcium	1.20	2.23	1.26	1.07	1.84

Crude protein (kjedahl), Crude fiber (ANKOM Fiber Analyzer) and Calcium (Flame AAS @ F.A.S.T. Laboratories, Mandaue, Cebu City.

Ash (ignition@600 celsius) and Crude fat (Soxhlet)@ Feed Lab, DA Region 7, Guadalupe, Mandaue Cebu City

Table 2. Average Number of Eggs Produced by Quail Birds Fed with Fresh *Trichanthera gigantea* and *Moringa oleifera* leaves.

Weeks	Treatments						P-value
	T0	T1	T2	T3	T4	T5	
Week 2	10.430 <sup>a</sup>	3.100 <sup>bc</sup>	2.167 <sup>bc</sup>	7.430 <sup>ab</sup>	5.730 <sup>abc</sup>	1.429 <sup>c</sup>	0.000
Week 4	13.571 <sup>a</sup>	4.643 <sup>bc</sup>	1.692 <sup>d</sup>	5.214 <sup>bc</sup>	6.000 <sup>b</sup>	3.214 <sup>cd</sup>	0.000
Week 6	15.587 <sup>a</sup>	7.692 <sup>b</sup>	2.846 <sup>c</sup>	8.929 <sup>b</sup>	7.000 <sup>b</sup>	6.417 <sup>b</sup>	0.000
Week 8	14.429 <sup>a</sup>	6.643 <sup>b</sup>	2.929 <sup>c</sup>	8.071 <sup>b</sup>	6.143 <sup>b</sup>	6.214 <sup>b</sup>	0.000

<sup>abcd</sup>Means with different letters indicate significant differences using Tukey's test ( $P$ -value<0.05).

the lowest, and there were slight differences among T1, T3, T4 and T5 which is the same as reported by Shen *et al.* (2021) at 10% of *M. oleifera* leaf meal feed inclusion for laying chicken. This result is somewhat different from those of previous studies (Abou-Ellez *et al.*, 2011; Abou-Ellez *et al.*, 2012; Lu *et al.*, 2016; Chen *et al.*, 2020; N'Nanle *et al.*, 2020), which found that high levels of *M. oleifera* leaf powder (MOLP) reduced laying performance, whereas low levels of MOLP did not. The results of MOLP supplementation may depend on the breed of chicken. Moreover, the protein digestibility of *M. oleifera* leaf was about 41.42% in the study of Alain *et al.* (2016) which was lower than soybean meal by 60% in the study of Saki *et al.* (2009). The results revealed that all levels of *M. oleifera* seed meal (MOSM) had recorded higher body weight (BW) and body weight gain (BWG) compared to the control group, and 7.5% inclusion of MOSM had the best result. During the overall period, chicks fed 5% MOSM recorded significantly lower feed conversion (FC) than the control (Rory *et al.*, 2016).

Replacement of 6% *M. oleifera* leaf meal given to Rhode Island chicks produced a significant increase in the feed intake, crude protein intake, average weight gain and feed efficiency compared to the control, which is contrary to the present study and these might also influence breed of animals (Melesse *et al.*, 2011). Bandura *et al.* (2020) reported that the supplementation of 4-6% MOLP in diets increased egg production in the laying hens, in contrast to the present study that shows the lower egg production of Japanese quail. Birds fed with 15% *T. gigantea* leaf in their feed have a delayed point of lay than those in control. Bejar (2017) also added that 15% inclusion of *T. gigantea* leaf meal is the maximum level beneficial for the growth, production and egg quality improvement of quail.

Furthermore, there were no significant differences in the average body weight of Japanese quail during week 2 and 4 and significant differences on week 6 and 8 among treatments. Treatment 2, fed with 30g commercial feeds and 10g fresh *M. oleifera*, has the lowest body weight which is also correlated with the feed consumption, as shown in Tables 3 and 4. Moreover, there were no significant differences on week 2 among treatments on the average feed consumption of Japanese quail, but significant differences were noticeable on week 4 to 10. As shown in Table 3, T2 was the lowest compared among the treatments in terms of average body weight and feed consumption.

Moreover, Ayssiwede *et al.* (2011) reported that the use of 24% *M. oleifera* leaves in the ration of indigenous Senegal chicken decreased feed intake, similar to the result of the present study as shown in Table 3. It was also as stated by Voemesse *et al.* (2018) that chicks fed a diet containing 3% *M. oleifera* showed a significantly heavier final weight similar to

the result of T3, T4 and T5 of Japanese quail as shown in Table 4. The effect of supplementation is that *M. oleifera* leaves ascription of 15% and 20% in the diet of poultry amassed body weight. In comparison, the ascription of 5% and 10% in poultry diet of broilers had no visible effect on the body weight, as also reported by Alnidawi *et al.* (2016). Forages having high pH levels, such as *T. gigantea* and Kakawate, had significantly higher voluntary intakes of organic matter (OM) and neutral detergent fiber (NDF) than forages with medium and low pH contents, such as santol, robles, gmelina and acacia (Aban *et al.*, 2015).

Alabi *et al.* (2017) verified that broilers fed diets containing aqueous *M. oleifera* leaf extracts had increased body weight, low feed intake and improved feed conversion ratio compared with the pure commercial feed or the control group. These results might be related to different bioactive constituents and nutrient exploitation in *M. oleifera* leaf extracts. The pharmacological chemical compounds (carbohydrates, saponins, cardiac glycosides, terpenes, steroids, flavonoids and alkaloids) are present in the extract as reported by Ambali and Furo (2012). This result is similar to the report of Zanu *et al.* (2012) who observed that the final body weight (FBW) and daily body weight gain (DBWG) increased with an increase in dosage level until 10% and then significantly declined with increasing levels of *M. oleifera* leaf meal. Unlike the results from previous researchers (Ashong and Brown, 2011; Portugaliza and Fernandez, 2012; Cassius Moreki and Gabanakgosi, 2014), birds on the aqueous *M. oleifera* extracts (AMOLE) treatments had significantly ( $P < 0.05$ ) depressed or lowered the FBW than the control. This might be because *M. oleifera* leaf meal was incorporated into the experimental diet instead of the aqueous extracts used in the

Table 3. Average Feed Consumption of Quail Birds Fed with Fresh *Trichanthera gigantea* and *Moringa oleifera* leaves.

Weeks	Treatments						P-value
	T0	T1	T2	T3	T4	T5	
Week 2, g	498.20	529.90	460.80	533.50	526.60	524.10	0.797
Week 4, g	390.71 <sup>a</sup>	387.79 <sup>a</sup>	373.39 <sup>c</sup>	384.79 <sup>ab</sup>	383.43 <sup>ab</sup>	379.29 <sup>bc</sup>	0.000
Week 6, g	391.07 <sup>a</sup>	387.79 <sup>a</sup>	363.21 <sup>c</sup>	385.50 <sup>ab</sup>	376.21 <sup>b</sup>	385.43 <sup>ab</sup>	0.000
Week 8, g	392.09 <sup>a</sup>	387.60 <sup>a</sup>	359.73 <sup>c</sup>	384.09 <sup>ab</sup>	385.00 <sup>b</sup>	384.09 <sup>ab</sup>	0.000

<sup>abc</sup>Means with different letters indicate significant differences using Tukey's test ( $P$ -value $<0.05$ ).

Table 4. Average Mean Value Body Weight of Quail Birds Fed with Fresh *Trichanthera gigantea* and *Moringa oleifera* leaves.

Weeks	Treatments						P-value
	T0	T1	T2	T3	T4	T5	
Week 2, g	140.60	133.85	131.48	132.36	137.55	135.55	0.024
Week 4, g	142.91	142.55	137.44	138.90	137.07	136.92	0.465
Week 6, g	151.56 <sup>abc</sup>	146.76 <sup>bc</sup>	145.59 <sup>c</sup>	145.25 <sup>c</sup>	157.36 <sup>ab</sup>	160.15 <sup>a</sup>	0.000
Week 8, g	164.04 <sup>a</sup>	161.08 <sup>a</sup>	148.17 <sup>c</sup>	155.22 <sup>ab</sup>	156.60 <sup>ab</sup>	156.38 <sup>ab</sup>	0.009

<sup>abc</sup>Means with different letters indicate significant differences using Tukey's test ( $P$ -value $<0.05$ ).

present study. The antimicrobial (lipophilic compounds) and antioxidant (polyphenols, tannins, anthocyanin, glycosides compound) present in *M. oleifera* leaf extracts (MOLE) may attach to the cytoplasmic membrane and remove free radicals, activate antioxidant enzymes and inhibit oxidases, thus, making these elements more available for the birds to use (Jabeen *et al.*, 2008; Luqman *et al.*, 2012). Furthermore, the synergy between individual bioactive compounds in *M. oleifera* leaf meal (MOLM) extract may be an essential feature of their action. It may affect broad aspects of physiology, such as nutrient absorption and processing, redox state, or immunity (Wallace *et al.*, 2010; Mikey, 2012).

Meanwhile, Abou-Ellez *et al.* (2011) confirmed undesirable effect wherein they detected decreased linear relation between the *M. oleifera* leaf dosage and egg-laying rate. This result may be due to the type of basal diets and the age of the harvested *M. oleifera*. As leaf age grows, crude fiber content becomes higher, and this increase may distress feed intake. The average daily gain was not significantly affected by varying levels of *T. gigantea* supplementation, slightly comparable to the result in the present study (Morboos *et al.*, 2016).

There were no significant differences in egg weight, albumen weight, yolk weight, egg breadth, and egg length between the treatments, as shown in Table 5. Egg weight is a trait influenced by genetics and protein in rations. Based on Table 5, it appears the egg weight varies among treatments from week 2 until week 8. There are significant differences during weeks two and four, but on week 8, there is no significant difference among treatments. The egg weight ranges from 9.041g to 10.603g. The result shows T5 with 3g fresh *T. gigantea* and 7g fresh *M. oleifera* leaves have an increasing trend throughout the week.

Nevertheless, there were significant differences in both albumen and yolk weight in the last week 6 among treatments. Eggs with the heaviest yolks and the largest yolk and albumen (Y:A) ratio would likely contain the highest cholesterol. Furthermore, there were significant differences in egg breadth during weeks 2 and 4 among treatments. Furthermore, on week 6, no significant differences were noticed. However, Table 5 shows that among the treatments, T2 had the widest eggs from weeks 2 to 6. Moreover, the egg length revealed no significant differences, but T0 and T5 indicate an increasing trend.

However, significant differences were observed in eggshell thickness at week 6 wherein T5 had the highest eggshell thickness among the treatments while T3 had the lowest eggshell thickness. This might influence the absorption of the mineral's nutrients in the Japanese quail birds. Nevertheless, the amount of ash represents the content level of total minerals. *M. oleifera* is rich in mineral elements, such as calcium, iron, potassium, phosphorus, and zinc, the key elements for animal growth and development (Teixeira *et al.*, 2014). Bardos *et al.* (2019) reported no differences in the quality and composition of eggs between improved Philippine Mallard Duck (IPMD) on the inclusion of *T. gigantea*; the finding indicates that there is no influence on the quality of yolk, albumen, and eggshell, irrespective of the feeding level and *T. gigantea* was not a factor for yolk color. *T. gigantea* had a moderate crude protein and a very high amount of calcium. As reported by Bidura *et al.* (2020), the effects of dietary supplementation with *Moringa* leaves powder on the external quality characteristics of laying hens exhibited higher significantly different ( $P < 0.05$ ) in eggshell thickness in the inclusion of 4% and 5% contrary to the present study. Furthermore, (Rory *et al.*, 2016) revealed the albumin and albumin/globulin (A/G) ratio was significantly decreased in birds fed 7.5% MOSM compared to the control. Sharmin *et al.* (2021) observed that the egg weight, length, width, shape index, and shell thickness of the eggs laid by hens fed diets with *M. oleifera* were similar during the experimental period.

Table 5. Egg quality of Japanese quail birds fed different treatment diets.

Weeks	Treatments	Parameters					
		Egg Weight, g	Egg Breadth, mm	Egg Length, mm	Egg Shell Thickness, mm	Albumen Weight, g	Yolk Weight, g
Week 2	T0	9.031 <sup>b</sup>	22.613 <sup>b</sup>	29.220	0.249 <sup>a</sup>	4.316	2.802 <sup>b</sup>
	T1	9.430 <sup>b</sup>	24.093 <sup>ab</sup>	29.427	0.159 <sup>b</sup>	4.675	3.067 <sup>b</sup>
	T2	10.603 <sup>a</sup>	25.400 <sup>a</sup>	30.013	0.230 <sup>a</sup>	4.716	3.667 <sup>a</sup>
	T3	9.526 <sup>b</sup>	24.100 <sup>ab</sup>	29.900	0.233 <sup>a</sup>	4.613	3.164 <sup>a</sup>
	T4	9.544 <sup>b</sup>	24.120 <sup>ab</sup>	29.833	0.265 <sup>a</sup>	4.587	3.138 <sup>b</sup>
	T5	9.422 <sup>b</sup>	23.947 <sup>ab</sup>	29.607	0.281 <sup>a</sup>	4.390	3.277 <sup>ab</sup>
	<i>P</i> -value	0.000	0.002	0.278	0.000	0.263	0.000
Week 4	T0	9.603 <sup>ab</sup>	23.893 <sup>b</sup>	30.427	0.267 <sup>ab</sup>	4.753 <sup>ab</sup>	3.036
	T1	10.034 <sup>a</sup>	24.680 <sup>a</sup>	30.073	0.281 <sup>a</sup>	5.185 <sup>a</sup>	3.212
	T2	9.695 <sup>ab</sup>	24.160 <sup>ab</sup>	29.873	0.306 <sup>a</sup>	5.047 <sup>ab</sup>	3.078
	T3	9.240 <sup>ab</sup>	23.720 <sup>b</sup>	29.893	0.230 <sup>b</sup>	4.785 <sup>ab</sup>	2.821
	T4	9.231 <sup>b</sup>	23.833 <sup>b</sup>	29.100	0.286 <sup>a</sup>	4.891 <sup>ab</sup>	2.899
	T5	9.589 <sup>ab</sup>	24.073 <sup>ab</sup>	29.853	0.287 <sup>a</sup>	4.575 <sup>b</sup>	3.059
	<i>P</i> -value	0.040	0.003	0.144	0.000	0.025	0.075
Week 6	T0	9.952	24.093	30.663	0.311 <sup>b</sup>	5.810	3.225
	T1	9.721	24.307	29.360	0.297 <sup>bc</sup>	4.863	3.047
	T2	9.917	24.433	29.720	0.314 <sup>b</sup>	4.982	3.293
	T3	9.761	24.093	30.173	0.247 <sup>c</sup>	5.101	3.029
	T4	9.501	24.013	29.487	0.331 <sup>ab</sup>	4.737	3.085
	T5	9.921	24.073	30.507	0.376 <sup>a</sup>	4.930	3.154
	<i>P</i> -value	0.591	0.479	0.018	0.000	0.224	0.321

<sup>abc</sup>Means with different letters indicate significant differences using Tukey's test (*P*-value<0.05).



These indicate that feeding with *Moringa* leaf meal up to 1.5% had no adverse effects on the eggs' external or internal qualities, similar to the present study. In contrast to the eggshell thickness, this indicates that the inclusion of 3g of *T. gigantea* and 7g of *M. oleifera* leaf have a capacity for better eggshell thickness in the long run.

The egg production was higher, while egg weight was heavier, yolk color was darker, and the FCR was better in layer quail supplemented with 5% *Moringa* leaf meal ( $P<0.05$ ) as well as in White Leghorns provided with 100 mL *Moringa* leaves extract added to 1Li of water ( $P<0.05$ ). On the other hand, the supplementation of *Moringa* seed meal in the diet of Babcock layers reduced the percentage of broken eggs (0.017 vs. 0.032;  $P<0.05$ ). Meanwhile, the ad libitum feeding of fresh *Moringa* leaves to broiler chickens did not affect body weight gain (Briones *et al.*, 2015).

The cost-benefit ratio is typically used for cost-benefit analyses, along with other measures such as the net present value, return on investment, internal rate of return, etc. Considering absolute amounts of cost and benefits sets this ratio apart from many other indicators. Table 6 shows the cost-benefit ratio analysis wherein among the treatments, T0 is better than treated diest but T3 and T4 also show potential in 5 years projection. This is indicated by a value of 1.20 and 1.21 wherein a value of more than 1.0 has a capacity to gain in the long run of production.

The results obtained in this study revealed that the inclusion of different percentages of fresh *M. oleifera* and *T. gigantea* in quail birds did not affect the production performance in terms of egg number and egg quality. However, on external quality, eggshell thickness in 3g *T. gigantea* and 7g *M. oleifera* (T5) have better results and show potential for 5 years of utilization. Therefore, the said inclusion rate of fresh *M. oleifera* and *T. gigantea* leaves could be utilized in quail layer diets. Furthermore, follow-up research is required to know the mechanism on the effects of the treatments, especially on the hatchability processes of the eggs.

Table 6. Cost-benefit ratio analysis of the different treatments.

Treatments	Cost-Benefit Ratio
T0	1.78
T1	0.93
T2	0.42
T3	1.20
T4	1.21
T5	0.77

## REFERENCES

- Aban ML, Gonzales LMR and Bestil LC. 2015. Voluntary feed intake in goats of foliage's with varying pH levels from selected trees and shrubs. *Int J Sci Res Publ* 5(2):1-4.
- Abbas TE. 2013. The use of *Moringa oleifera* in poultry diets. *Turk J Vet Anim Sci* 37(5):492-496.

- Abou-Ellez, Sarmiento FL, Santos RR and Solorio SF. 2011. Nutritional effects of dietary inclusion of *Leucaena leucocephala* and *Moringa oleifera* leaf meal on Rhode Island Red hen's performance. *Cuban J Agric Sci* 45:163-9.
- Abou-Ellez, Sarmiento FL, Santos RR and Solorio JF. 2012. The nutritional effect of *Moringa oleifera* fresh leaves as feed supplement on Rhode Island Red hen egg production and quality. *Trop Anim Health Prod* 44(5):1035–1040.
- Alabi OJ, Malik AD, Ng'Ambi JW, Obaje P and Ojo BK. 2017. Effect of aqueous *Moringa oleifera* (Lam) leaf extracts on growth performance and carcass characteristics of Hubbard broiler chicken. *Braz J Poult Sci* 19:273-280.
- Alain MM, Nyobe EC, Bakwo BC and Minka SR. 2016. A comparison of the nutritional quality of proteins from *Moringa oleifera* leaves and seeds. *Cogent Food Agric* 2(1):1213618.
- Alnidawi A, Ali F, Abdelgayed S, Ahmed F and Farid M. 2016. *Moringa oleifera* leaves in broiler diets: Effect on chicken performance and health. *Food Sci Qual Manage* 58:40-48.
- Ambali AG and Furo NA. 2012. An investigation into the phytochemical constituents of *Moringa oleifera* aqueous root extracts. *Thesis*. Maiduguri (NIG): Faculty of Veterinary Medicine, University of Maiduguri.
- Ashong JO and Brown DL. 2011. Safety and efficacy of *Moringa oleifera* powder for growing poultry. *J Anim Sci* 89:84.
- Ayssiwede SB, Dieng A, Bello H, Chrysostome CM, Hane M B, Mankor A, Dahouda M, Houinato MR, Hornick JL and Missohou A. 2011. Effects of *Moringa oleifera* (Lam.) leaves meal incorporation in diets on growth performances, carcass characteristics and economics results of growing indigenous Senegal chickens. *Pak J Nutr* 10(12):1132-45
- Bahadori Z, Esmailzadeha L, Karimi-Torshizib MA, Seidavic A, Olivarese J, Rojas S, Saleme AZM, Khusrof A and Lópezg S. 2017. The effect of earthworm (*Eisenia foetida*) meal with vermi-humus on growth performance, hematology, immunity, intestinal microbiota, carcass characteristics, and meat quality of broiler chickens. *Livest Sci* 202:74-81.
- Bejar FR. 2017. Madre de Agua (*Trichanthera gigantea*) Leaf Meal as Fed to Quails with Aloe Vera Extract and Acid Cheese Whey Supplementation. *Countryside Dev Res J* 5(01):15-23.
- Berdos JL, Martin EA, Celestino OF, Paragas and Ella JS. 2019. Egg Production Performance of Improved Philippine Mallard Ducks (*Anas platyrhynchos*) fed diets supplemented with fresh trichanthera (*Trichanthera gigantea*) leaves. *Phil J Vet Anim Sci* 201945(1):48-57.
- Bidura IGNG, Partama IBG, Utami IAP, Cnadrawati DPMA, Puspani E, Suasta IM, Warmadewi DA, Okarini IA, Wibawa AAP, Nuriyasa IM and Siti NW. 2020. *Effect of Moringa oleifera leaf powder in diets on laying hen's performance, b-carotene, cholesterol, and minerals contents in egg yolk*. IOP Conference Series: Materials Science and Engineering, Kupang, Indonesia.
- Briones J, Leung A, Bautista N, Golin S, Caliwag N, Carlos MA and Mendoza D. 2017. Utilization of *Moringa oleifera* Lam. in animal production. *Acta Horti* 1158:467-474.
- Cassius Moreki J and Gabanakgosi K. 2014. Potential use of *Moringa oleifera* in poultry diets. *Glob J Anim Sci Res* 2-2(2):109-115.

- Chen ZM, Chang WH, Zheng AJ, Cai HY and Liu GH. 2020. Tolerance evaluation of *Moringa oleifera* extract to Hailan brown laying hens. *J Anim Physiol Anim Nutr* 104(5):1375–1379.
- Genchev A. 2012. Quality and composition of Japanese quail eggs (*Coturnix japonica*). *Trakia J Sci* 10(2):91-101.
- Hasan M, Sharmeen IA, Anwar Y, Alharby HF, Hasanuzzaman M, Hajar AS and Hakeem KR. 2019. Evidence-based assessment of *Moringa oleifera* used for the treatment of human ailments. In *Plant and Human Health* 2:121-137. DOI: [https://doi.org/10.1007/978-3-030-03344-6\\_4](https://doi.org/10.1007/978-3-030-03344-6_4).
- Hien TQ, Hoan TT, KHOA M, Kien TT and Trung TQ. 2018. Comparison of the Effects of Several Leaf Meal Kinds Included in Diets of Laying Hens on Egg Yield and Quality. *Bulg J Agric Sci* 24(2):303-309.
- Jabeen R, Shahid M, Jamil A and Ashraf M. 2008. Microscopic evaluation of the antimicrobial activity of seed extracts of *Moringa oleifera*. *Pak J Bot* 40:1349–1358.
- Lalwani P. 2011. *Quail egg nutrition*. Retrieved on 25 August 2021 from <http://www.buzzle.com/articles/quailegg-nutrition.html>.
- Lu W, Wang J, Zhang HJ, Wu SG and Qi GH. 2016. Evaluation of *Moringa oleifera* leaf in laying hens: effects on laying performance, egg quality, plasma biochemistry and organ histopathological indices. *Ital J Anim Sci* 15(4):658–665.
- Luqman S, Srivastava S, Kumar R, Maurya AK and Chanda D. 2012. Experimental assessment of *Moringa oleifera* leaf and fruit for its antistress, antioxidant, and scavenging potential using in vitro and in vivo assays. *Evid Based Complement Alternat Med* DOI: <https://doi.org/10.1155/2012/519084>
- Malkanathi SHP, Dilini Rathnachandra SD and Weerasinghe WARN. 2021. Consumers' Awareness on Organic Food: Case of Urban Sri Lanka. *Sci J Wars Univ Life Sci-SGGW* 21(4):25-36.
- Melesse A, Tiruneh W and Negesse T. 2011. Effects of feeding *Moringa stenopetala* leaf meal on nutrient intake and growth performance of Rhode Island Red chicks under tropical climate. *Trop Subtrop Agroecosystems* 14:485-492.
- Mikey M. 2012. Therapeutic potential of *Moringa oleifera* leaves in chronic hyperglycemia and dyslipidemia: A Review. *Front Pharmacol* 3(24):1-12.
- Morbos CE, Espina DM and Bestil LC. 2016. Growth Performance of Philippine Native Chicken Fed Diet Supplemented with Varying Levels of Madre de Agua (*Trichanthera gigantea* Nees) Leaf Meal. *Ann Trop Res* 38(1):174-182.
- N'Nanle O, Tété BA, Nideou D, Onagbesan OM and Tona K. 2020. Use of *Moringa oleifera* leaves in broiler production chain. Effect on Sasso breeder hens' performances, internal quality of hatching eggs and serum lipids. *Vet Med Sci* 6(3):485–490.
- Offiong AA, Nduononwi AA, Edet BL and Isonguyo ES. 2020. *Technical Vocational Education and Training (TVET) Opportunities in Quail Production for Gainful Self-Employment Among In-College and Out-of-College Youths in Nigeria*. World Journal of Interactive Research 2(1). Retrieved on 21 October 2021 from <https://benchmarkjournals.com>.
- Oladipo GO and Ibukun EO. 2017. Bio Activities of *Coturnix japonica* (quail) egg yolk and albumen against physiological stress. *Food Sci Nut* 5(2):334-343.

- Olorunfemi OD, Oladipo FO, Bolarin O, Akangbe JA and Bello OG. 2016. Capacity building needs of poultry farmers for quail production in Kwara State, Nigeria. *J Agric Sci (Belgr)* 61(1):69-78.
- Portugaliza HP and Fernandez TJ. 2012. Growth performance of Cobb broilers given varying concentrations of malunggay (*Moringa oleifera* Lam.) aqueous leaf extract. *J Anim and Feed Res* 2(6):465-469.
- Pramono A, Primadhani MS, Swastike W and Sutrisno J. 2018. Nutrient digestibility of vegetables waste flour on male quail (*Coturnix coturnix japonica*). *IOP Conf. Series: Earth Environ Sci* 142 DOI:10.1088/1755-1315/142/1/012025.
- Ronald PC and Adamchak RW. 2018. *Tomorrow's table: organic farming, genetics and the future of food*. Oxford University Press: United State of America.
- Rory FH, Elkloub K, Moustafa, MEL, Mousa MAM, Youssef SF and Hanan AH. 2016. *Effect of using Moringa oleifera seed meal on Japanese quail performance during growing period*. In Proceedings of the 9th Intl Poult Con, Hurghada, Egypt.
- Saki AA, Abbasinezhad M, Ghazi S, Tabatabai MM, Goodarzi M, Ahmadi A, Zaboli K, Karami O, Zamani, Z and Mahmoudi H. 2009. Protein pattern and urease activity of two types of soybean meal on protein digestibility and chicken performance. *J Appl Anim Res* 35(1):45-48.
- Sharmin F, Karim S, Sazedul MD, Sarker NR and Faruque S. 2021. Dietary effect of *Moringa oleifera* on native laying hens egg quality, cholesterol and fatty – acid profile. *Ital J Anim Sci* 20:1:1544-1553.
- Shen M, Li T, Qu L, Wang K, Hou Q, Zhao W and Wu P. 2021. Effect of dietary inclusion of *Moringa oleifera* leaf on productive performance, egg quality, antioxidant capacity and lipid levels in laying chickens. *Ital J Anim Sci* 20(1):2012-2021.
- Teixeira EMB, Carvalho MRB, Neves VA, Silva MA and Arantes-Pereira L. 2014. Chemical characteristics and fractionation of proteins from *Moringa oleifera* Lam. leaves. *Food Chem* 147:51-54.
- Tolik D, Poawska E, Charuta A, Nowaczewski S and Cooper R. 2014. Characteristics of egg parts, chemical composition and nutritive value of Japanese quail eggs—a review. *Folia Biol* 62(4):287-292.
- Tunsaringkarn T, Tungjaroenchai W and Siriwong W. 2013. Nutrient benefits of quail (*Coturnix coturnix japonica*) eggs. *Int J Sci and Res Publ* 3(5):1-8.
- Voemesse K, Teteh A, Nideou D, Nanle OM, Gbeassor M, Decuypere E and Tona K. 2018. Effect of *Moringa oleifera* leaf meal on Growth Performance and Blood Parameters of Egg Type Chicken during Juvenile Growth. *Intl J Poult Sci* 17: 154-159.
- Wallace RJ, Oleszek W, Franz C, Hahn I, Baser KHC and Mathe A. 2010. Dietary plant bioactives for poultry health and productivity. *Br Poult Sci* 51(4):461-487.
- Wilson R and Hettinger R. 2017. *Food and Nutrition Research Institute (FNRI), Philippines*. Retrieved on 12 October 2021 from [https://digitalcommons.iwu.edu/freeman\\_posters/15](https://digitalcommons.iwu.edu/freeman_posters/15).
- Zanu HK, Asiedu P, Tampuori M, Abada M and Asante I. 2012. Possibilities of using *Moringa (Moringa oleifera)* leaf meal as a partial substitution for fishmeal in broiler chicken diets. *J Anim Feed Res* 2(1)70-75.